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Editorial

*Corresponding author Juan Sanchez-Esteban, MD Associate Professor Department of Pediatrics Women and Infants Hospital of Rhode Island The Warren Alpert Medical School of Brown University 101 Dudley Street. Providence Rhode Island 02905, USA Tel. 401-274-1122 Fax: 401-453-7571 E-mail: jsanchezesteban@wihri.org

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Alveolar Type I Epithelial Cells: The Forgotten Cells in Fetal Lung Development and Lung Injury

Tanbir Najrana and Juan Sanchez-Esteban*

Department of Pediatrics, Women and Infants Hospital of Rhode Island and The Warren Alpert Medical School of Brown University, Providence, Rhode Island 02905, USA

The alveolar surface of the lung is covered by large flat type I epithelial cells. Even though type I cells represent only around 10% of the cells present in the alveolus; they cover much of the surface area in the developed lung.¹ Given their thinness and proximity to the capillary endothelium; it is well accepted that type I cells play an important role in gas exchange.² In addition, these cells are important to maintain adequate fluid balance in the alveolus³ *via* the tight junctions,⁴ ion transport channels⁵ and aquaporin-5.⁶ Recent studies also indicate that type I cells participate in innate immunity; they express toll-like receptor 4 and produce pro-inflammatory cytokines.^{7,8} Studies from T1 α knockout mice indicate that alveolar type I cells may be critical for normal lung development. T1 α , a lung type I cell differentiation gene, is developmentally regulated and expressed only in type I cells. T1 α knockout mice died at birth of respiratory failure. Histologic analysis show fewer alveolar type I cells and decreased alveoli.⁹ All together, these investigations suggest a critical role for type I cells in gas exchange, alveolar fluid hemostasis, immunity and fetal lung development.

The typical flat morphology of type I cells begin to appear in the late canalicular period and increase in number during the saccular and alveolar stages of lung development.¹⁰ It has been believed that type I cells are derived from type II cells.^{11,12} However, recent studies¹³ using specific markers for type I (T1alpha (T1a) and Receptor for Advanced Glycation Endproducts (RAGE))) and type II cells (SP-C, NKX2-1, and ABCA3) have demonstrated the presence in the distal lung of alveolar progenitor cells containing both phenotypes, before they became differentiated type I or type II cells. Therefore, these studies show that during fetal lung development, alveolar type I and type II epithelial cells are derived from a bipotent progenitor cell.¹³ Hooper's group found that the numbers of "intermediate cells" expressing both phenotypes were strongly influenced by the degree of lung expansion,¹⁴ supporting the role of mechanical signals in fetal lung development and differentiation of alveolar epithelial cells.

Many premature infants born with underdeveloped lungs develop Bronchopulmonary dysplasia (BPD), a chronic inflammatory lung disease with serious short- and long-term complications. Although the etiology of BPD is multifactorial, mechanical ventilation plays a central role.¹⁵ Excessive stretch of the lung by mechanical ventilation can disrupt the integrity of the alveolar-capillary barrier, resulting in interstitial and alveolar edema. Neutrophils and macrophages recruited to the lung can then trigger and amplify an injury response by releasing cytokines and other inflammatory mediators.^{16,17} Many of these pro-inflammatory cytokines are secreted by alveolar macrophages, fibroblasts, type II pneumocytes, and endothelial cells.¹⁸ Distal lung parenchyma cells can be directly exposed to overstretch, and therefore to injury secondary to mechanical ventilation. It has been shown for example that type II epithelial cells release proinflammatory cytokines in response to mechanical injury.¹⁹⁻²² Given that type I epithe lial cells cover much of the distal epithelium of the lung, these cells are also at risk for injury mediated by mechanical ventilation. However, the contribution of type I cells to the pathogenesis of BPD is not clearly defined, in part because of the difficulty in isolating type I cells in vitro.²³ Nevertheless, recent studies have found these cells produce Tumor Necrosis Factoralpha (TNF-α), Interleukin-1 beta or IL-1beta (IL-1β), and Interleukin 6 (IL-6) after exposure to Lipopolysaccharide (LPS).²⁴ In fact, some authors believe that alveolar type I epithelial cells

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are a more important source of pro-inflammatory cytokines than type II cells.²⁵ Moreover, the Receptor for advanced glycation endproducts (RAGE) is found only on type I cells in the lung.²⁶ RAGE signaling is mediated *via* NF-κB pathway, stimulating production of pro-inflammatory cytokines and inducing apoptosis.²⁷

The epithelial barrier is composed of tight junctions connected to the actin cytoskeleton *via* occludin or zonula occludens. It has been shown that mechanical strain of alveolar epithelial cells, mimicking mechanical ventilation with high tidal volumes, resulted in actin-mediated cell contraction with subsequent increased in paracellular permeability²⁸ and breakdown of intercellular junctions.^{29,30} These junctions could be affected by mechanical injury, leading to pulmonary edema.^{31,32} In addition to maintaining the integrity of the epithelial barrier by the tight junctions, epithelial cells need mechanisms to reabsorb the fluids present in the interstitium and alveolar spaces after lung injury mediated by mechanical ventilation.³³ This process is mediated by active transport of Na⁺ through amiloride-sensitive cation channels Epithelial Na⁺ Channels (ENaC) present in the apical cell membranes and the Na⁺/K⁺-ATPases localized mainly in the basolateral cell membrane.^{34,37} Electron microscope studies provided clear evidence for the major abnormalities in the blood-gas barrier during lung injury. Damage of alveolar type I epithelial cells was observed in rabbits ventilated with a peak inspiratory pressure of 20 cm H₂O for 6 hours.³⁸ In these studies, some endothelial cells were detached from their basement membrane, resulting in the formation of intra-capillary blebs. There were also occasional breaks in endothelial cells. More prolonged exposure to injurious stress produced alveolar epithelial pathology ranging from inter- and intra-cellular gap formations with denuded basement membranes to extensive cell destruction.³⁹

In summary, and as discussed in an excellent review by Dr. Rozycki,²³ alveolar development requires an orchestrated signaling cross-talk among different cells of the distal lung.⁴⁰ Given that type I epithelial cells are critical for normal lung development and to maintain the hemostasis of the distal lung, damage of these cells and/or their progenitors by mechanical ventilation and hyperoxia could not only disrupt normal pulmonary development but also have a significant contribution to the pulmonary edema and inflammation observed in patients with BPD. Future studies will provide more insights into the role of these forgotten cells in fetal lung development and lung injury of premature lungs.

CONFLICTS OF INTEREST

The authors declare that they have no conflicts of interest.

REFERENCES

1. Stone KC, Mercer RR, Gehr P, Stockstill B, Crapo JD. Allometric relationships of cell numbers and size in the mammalian lung. *Am J Respir Cell Mol Biol*. 1992; 6: 235-243. doi: 10.1165/ajrcmb/6.2.235

2. Makanya A, Anagnostopoulou A, Djonov V. Development and remodeling of the vertebrate blood-gas barrier. *Biomed Res Int.* 2013; 2013: 101597. doi: 10.1155/2013/101597

3. Johnson MD, Bao HF, Helms MN, et al. Functional ion channels in pulmonary alveolar type I cells support a role for type I cells in lung ion transport. *Proc Natl AcadSci USA*. 2006; 103: 4964-4969. doi: 10.1073/pnas.0600855103

4. Schneeberher EE, Lynch RD. The tight junction: a multifunctional complex. *Am J Physiol Cell Physiol*. 2004; 286: C1213-C1228. doi: 10.1152/ajpcell.00558.2003

5. Hollenhorst MI, Richter K, Fronius M. Ion transport by pulmonary epithelia. *J Biomed Biotechnol*.2011; 2011: 174306. doi: 10.1155/2011/174306

6. Ma T, Fukuda N, Song Y, Matthay MA, Verkman AS. Lung fluid transport in aquaporin-5 knockout mice. *J Clin Invest.* 2000; 105: 93-100.

7. Wong MH, Chapin OC, Johnson MD. LPS-stimulated cytokine production in type i cells is modulated by the renin-angiotensin system. *Am J Respir Cell Mol Biol.* 2012; 46: 641-650. doi: 10.1165/rcmb.2011-0289OC

8. Wong MH, Johnson MD. Differential response of primary alveolar type I and AEC2 cells to LPS stimulation. *PLoS One*. 2013; 8: e55545. doi: 10.1371/journal.pone.0055545

9. Ramirez MI, Millien G, Hinds A, Cao Y, Seldin DC, Williams MC. T1alpha, a lung type I cell differentiation gene, is required for

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http://dx.doi.org/10.17140/PRRMOJ-2-e003

normal lung cell proliferation and alveolus formation at birth. Dev Biol. 2003; 256: 62-73. doi: 10.1016/S0012-1606(02)00098-2

10. Flecknoe SJ, Wallace MJ, Cock ML, Harding R, Hooper SB. Changes in alveolar epithelial cell proportions during fetal and postnatal development in sheep. *Am J Physiol Lung Cell Mol Physiol*. 2003; 285: L664-L670. doi: 10.1152/ajplung.00306.2002

11. Evans MJ, Cabral LJ, Stephens RJ, Freeman G. Renewal of alveolar epithelium in the rat following exposure to NO₂. Am J Pathol. 1973; 70: 175-198.

12. Gabazza EC, Kasper M, Ohta K, et al. Decreased expression of aquaporin-5 in bleomycin-induced lung fibrosis in the mouse. *Pathol Int.* 2004; 54: 774-780.

13. Desai TJ, Brownfield DG, Krasnow MA. Alveolar progenitor and stem cells in lung development, renewal and cancer. *Nature*. 2014; 507: 190-194. doi: 10.1038/nature12930

14. Flecknoe SJ, Wallace MJ, Harding R, Hooper SB. Determination of alveolar epithelial cell phenotypes in fetal sheep: evidence for the involvement of basal lung expansion. *J Physiol*. 2002; 542: 245-253. doi: 10.1113/jphysiol.2001.014274

15. Tibboel D, Jobe AH. Update in pediatric lung disease 2009. *Am J Respir Crit Care Med.* 2010; 181(7): 661-665. doi: 10.1164/ rccm.201001-0117UP

16. Carlton DP, Albertine KH, Cho SC, Lont M, Bland RD. Role of neutrophils in lung vascular injury and edema after premature birth in lambs. *J Appl Physiol*. 1997; 83(4): 1307-1317.

17. Speer CP. Inflammation and bronchopulmonary dysplasia: a continuing story. *Seminars in Fetal & Neonatal Medicine*. 2006; 11(5): 354-362. doi: 10.1016/j.siny.2006.03.004

18. Speer CP. Inflammation and bronchopulmonary dysplasia. Semin Neonatol. 2003; 8(1): 29-38. doi: 10.1016/S1084-2756(02)00190-2

19. Hammerschmidt S, Kuhn H, Sack U, et al. Mechanical stretch alters alveolar type II cell mediator release toward a proinflammatory pattern. *Am J Respir Cell Mol Biol.* 2005; 33(2): 203-210. doi: 10.1165/rcmb.2005-0067OC

20. Lee HS, Wang Y, Maciejewski BS, et al. Interleukin-10 protects cultured fetal rat type II epithelial cells from injury induced by mechanical stretch. *Am J Physiol Lung Cell Mol Physiol*. 2008; 294(2): L225-L232. doi: 10.1152/ajplung.00370.2007

21. Vlahakis NE, Schroeder MA, Limper AH, Hubmayr RD. Stretch induces cytokine release by alveolar epithelial cells in vitro. *Am J Physiol*. 1999; 277(1 Pt 1): L167-L173.

22. Thorley AJ, Ford PA, Giembycz MA, Goldstraw P, Young A, Tetley TD. Differential regulation of cytokine release and leukocyte migration by lipopolysaccharide-stimulated primary human lung alveolar type II epithelial cells and macrophages. *J Immunol*. 2007; 178(1): 463-473. doi: 10.4049/jimmunol.178.1.463

23. Rozycki HJ. Potential contribution of type I alveolar epithelial cells to chronic neonatal lung disease. *Front Pediatr.* 2014; 2: 45. doi: 10.3389/fped.2014.00045

24. Wong MH, Chapin OC, Johnson MD. LPS-stimulated cytokine production in type I cells is modulated by the renin-angiotensin system. *Am J Respir Cell Mol Biol.* 2012; 46(5): 641-650.

25. Wong MH, Johnson MD. Differential response of primary alveolar type I and type II cells to LPS stimulation. *PLoS One*. 2013; 8(1): e55545.

26. Demling N, Ehrhardt C, Kasper M, Laue M, Knels L, Rieber EP. Promotion of cell adherence and spreading: a novel function of RAGE, the highly selective differentiation marker of human alveolar epithelial type I cells. *Cell Tissue Res.* 2006; 323: 475-488. doi: 10.1007/s00441-005-0069-0

27. Stogsdill JA, Stogsdill MP, Porter JL, Hancock JM, Robinson AB, Reynolds PR. Embryonic overexpression of receptors for



ISSN 2377-1658

= Open Journal 👌 =

http://dx.doi.org/10.17140/PRRMOJ-2-e003

advanced glycation end-products by alveolar epithelium induces an imbalance between proliferation and apoptosis. *Am J Respir Cell Mol Biol.* 2012; 47: 60-66. doi: 10.1165/rcmb.2011-0385OC

28. DiPaolo BC, Lenormand G, Fredberg JJ, Margulies SS. Stretch magnitude and frequency-dependent actin cytoskeleton remodeling in alveolar epithelia. *Am J Physiol Cell Physiol.* 2010; 299(2): C345-C353. doi: 10.1152/ajpcell.00379.2009

29. Garcia JG, Davis HW, Patterson CE. Regulation of endothelial cell gap formation and barrier dysfunction: role of myosin light chain phosphorylation. *J Cell Physiol.* 1995; 163(3): 510-522. doi: 10.1002/jcp.1041630311

30. Goeckeler ZM, Wysolmerski RB. Myosin light chain kinase-regulated endothelial cell contraction: the relationship between isometric tension, actin polymerization, and myosin phosphorylation. *J Cell Biol.* 1995; 130(3): 613-627.

31. Schneeberger EE, Lynch RD. The tight junction: a multifunctional complex. *Am J Physiol Cell Physiol*. 2004; 286(6): C1213-C1228. doi: 10.1152/ajpcell.00558.2003

32. Dipaolo BC, Davidovich N, Kazanietz MG, Margulies SS. Rac1 pathway mediates stretch response in pulmonary alveolar epithelial cells. *Am J Physiol Lung Cell Mol Physiol*. 2013; 305(2): L141-L153. doi: 10.1152/ajplung.00298.2012

33. Hochberg I, Abassi Z, Azzam ZS. Patterns of alveolar fluid clearance in heart failure. *Int J Cardiol.* 2008; 130(2): 125-130. doi: 10.1016/j.ijcard.2008.03.015

34. Goodman BE, Fleischer RS, Crandall ED. Evidence for active Na+ transport by cultured monolayers of pulmonary alveolar epithelial cells. *Am J Physiol*. 1983; 245(1): C78-C83.

35. Basset G, Bouchonnet F, Crone C, Saumon G. Potassium transport across rat alveolar epithelium: evidence for an apical Na⁺-K⁺ pump. *J Physiol.* 1988; 400: 529-543.

36. Matalon S, Benos DJ, Jackson RM. Biophysical and molecular properties of amiloride-inhibitable Na⁺ channels in alveolar epithelial cells. *Am J Physiol.* 1996; 271(1 Pt 1): L1-L22.

37. Sznajder JI, Olivera WG, Ridge KM, Rutschman DH. Mechanisms of lung liquid clearance during hyperoxia in isolated rat lungs. *Am J Respir Crit Care Med.* 1995; 151(5): 1519-1525. doi: 10.1164/ajrccm.151.5.7735609

38. John E, McDevitt M, Wilborn W, Cassady G. Ultrastructure of the lung after ventilation. Br J Exp Pathol. 1982; 63(4): 401-407.

39. Dreyfuss D, Basset G, Soler P, Saumon G. Intermittent positive-pressure hyperventilation with high inflation pressures produces pulmonary microvascular injury in rats. *Am Rev Respir Dis.* 1985; 132(4): 880-884.

40. Herriges M, Morrisey EE. Lung development: orchestrating the generation and regeneration of a complex organ. *Development*. 2014; 141: 502-513. doi: 10.1242/dev.098186